



Figure 2. ISH of carp embryos with carp *Hoxb-3* antisense RNA probe. (A) One somite stage embryo, dorsal view, posterior to the top. The black arrowheads point at the anterior bands of expression in the neurectoderm, the open arrowheads point at the posteriorly located bands in the longitudinal stripes of gene expression. (B) Longitudinal section of a one somite stage embryo. The black arrowhead points at anterior boundary of expression, the open arrowhead at one of the first pair of somites. (C) Double ISH on a 10 S stage embryo using *Hoxb-1* (red staining) and *Hoxb-3* (blue staining). The black arrowheads point at the *Hoxb-1* expression in r4, open arrowheads at the *Hoxb-3* expression in r5, r6, and r7. (D) Cross section through r6 or r7 at the 15 somites stage. Expression is especially strong in the ventral part of the brain, except for its most ventral aspect and the floorplate (arrow). The notochord (n) is also negative. The black arrowheads point at the *Hoxb-3* expression in the neural crest cells.

much stronger ventrally than dorsally, as can be seen in figure 2D. Also noticeable are the negative notochord, floorplate, and the layer of cells above the floorplate. Lateral from the neurale tube a paired group of cells was found that also expressed *Hoxb-3*. These cells most likely represent the neural crest. Sham *et al.* (1992) also found *Hoxb-3* expression in mouse neural crest cells. At 24h and 36h expression of *Hoxb-3* was also found in gill arches. Migrating neural crest cells may be involved.

To test the possibility to use the carp *Hoxb-3* probe in zebrafish, a cyprinid species closely related to carp (Stroband *et al.*, 1995), a whole mount ISH was carried out using 14 S stage zebrafish embryos. The expression patterns in carp and zebrafish were identical, but further studies of sections are needed.

In conclusion, the present study shows that homeobox sequences of *Hoxb-3* genes are very well conserved between fish and other vertebrates and even lower animals, and that its pattern of expression also seems to be conserved. However, concerning the latter, further study has to be done.

References

- Acampora D, D'Esposito M, Faiella A, Pannese M, Migliaccio E, Morelli F, Stornaiuolo A, Nigro V, Simeone A, and Boncinelli E. (1989) The human HOX gene family. *Nucleic Acids Res.* 17:10385-10402.
- Gehring WJ, Affolter M, Brglin T (1994) Homeodomain proteins. *Annu. Rev. Biochem.* 63:487-526.
- Godsave S, Dekker E-J, Holling T, Pannese M, Boncinelli E, Durston A (1994) Expression Patterns of *Hoxb* Genes in *Xenopus* Embryo Suggest roles in anteroposterior Specification of the Hindbrain and in Dorsoventral Patterning of the Mesoderm. *Dev. Biol.* 166:465-476.
- Misof BY, Blanco MJ, Wagner GP (1996) PCR-Survey of Hox-Genes of the zebrafish: New Sequence Information and Evolutionary Implications. *J. Exp. Zool.* 274: 193-206.
- Rex M and Scotting P J (1994) Chick HoxB3: deduced amino-acid sequence and embryonic gene expression. *Gene* 149:381-382.
- Sham MH, Hunt P, Nonchev S, Papalopulu N, Graham A, Boncinelli E, and Krumlauf R (1992) Analysis of the murine *Hox-2.7* gene: conserved alternative transcripts with differential distributions in the nervous system and the potential for shared regulatory regions. *EMBO J.* 11:1825-1836.
- Stevens CJM, Samallo J, Schipper H, Stroband HWJ, and Te Kronnie G (1996) Expression of *Hoxb-1* during gastrulation and segmentation stages of carp (*Cyprinus carpio*). *Int. J. Dev. Biol.* 40:463-470.
- Stroband H W J, Stevens C, Te Kronnie G, Samallo J, Schipper H, Kramer B, and Timmermans L P M (1995) Expression of *carp-cdx1*, a caudal homolog, in embryos of the carp, *Cyprinus carpio*. *Roux's Arch. Dev. Biol.* 204:369-377.