

Uterine natural killer cells as modulators of the maternal-fetal vasculature

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ABSTRACT Precise and local control of the innate immune system within the placenta is an essential component for achieving a normal and healthy pregnancy. One of the most abundant immune cells of the placenta is a subpopulation of natural killer (NK) cells that profusely populates the uterine decidua during early pregnancy. Uterine NK (uNK) cells and trophoblast cells of the placenta communicate both directly and indirectly to contribute to the critical process of spiral artery remodeling. Here, we discuss recent findings that expand our knowledge of uNK cell-trophoblast cell crosstalk and the important role it plays in the maternal vascular adaptation to pregnancy.

KEY WORDS: *NK cell, trophoblast cell, placenta, spiral artery remodeling, preeclampsia*

Introduction

Natural killer (NK) cells are lymphocytes belonging to the innate immune system that attack virally-infected cells and tumor cells via exocytosis of perforin- and granzyme-containing granules. In females, there are two populations of NK cells: peripheral blood (pb) NK cells and decidual (d) or uterine (u) NK cells. Both NK cell populations are capable of cytotoxicity and cytokine secretion. However, pbNK cells are primarily lytic cells, while uNK cells are primarily cytokine and chemokine producers.

A dramatic expansion of uNK cells occurs during early pregnancy, populating two adjacent areas of the implantation site, the decidua basalis (DB) and the mesometrial lymphoid aggregate of pregnancy (MLAp). Proliferation continues until mid-pregnancy, at which point uNK cells comprise up to 70% of immune cells present in the decidua, the progesterone-altered endometrium of the uterus that supports the conceptus (Bulmer *et al.*, 2010). uNK cell population size then declines until the end of pregnancy.

Molecular characterization of uNK cells has led to the identification of subpopulations of uNK cells in the decidua. Mouse uNK cells, previously identified by periodic acid Schiff (PAS) staining, are also currently identified by *Dolichos biflorus* agglutinin (DBA) lectin staining (Paffaro *et al.*, 2003). PAS and DBA lectin staining defines two subpopulations of uNK cells: PAS⁺DBA⁻ and PAS⁺DBA⁺ cells, which exhibit different gene expression profiles (Chen *et al.*, 2012, Zhang *et al.*, 2011). While PAS is a pan-uNK cell marker, DBA lectin detects the subpopulation of cells that expands during pregnancy, as 90% of uNK cells at mid-gestation are DBA⁺ (Zhang *et al.*, 2011).

In contrast to the mouse, human uNK cells are identified by their CD56⁺CD16⁻ signature. The vast majority of uNK cells are CD56^{bright}CD16⁻, and pbNK cells are typically CD56^{dim}CD16⁺. Interestingly, while both subpopulations are present in the decidua, the proportion of these two subsets can shift to favor cytotoxic CD56^{dim}CD16⁺ cells in the presence of infectious agents like cytomegalovirus (Siewiera *et al.*, 2013) and *Toxoplasma gondii* (Xu *et al.*, 2013a), both common intrauterine infections that cause severe birth defects.

Despite belonging to the immune system, uNK cells' primary contributions to the developing pregnancy are not immune in nature. Rather, uNK cell-secreted cytokines and chemokines communicate with fetal trophoblast cells of the placenta (Hanna *et al.*, 2006); these two cell types act in concert to remodel spiral arteries, conduits of blood from the uterus to the placental bed and growing fetus (Smith *et al.*, 2009). The importance of this process is stressed by the association of insufficient spiral artery remodeling with several diseases of pregnancy, such as fetal growth restriction (FGR) and preeclampsia. Here, we briefly review aspects of uNK cell-trophoblast cell crosstalk and their role in spiral artery remodeling and the maintenance of pregnancy, as summarized in Fig. 1.

Signals promoting differentiation of uNK cells

Little is known about uNK cell precursors and the source of the expanded uNK cell population during pregnancy. However, there

Abbreviations used in this paper: AM, adrenomedullin; DBA, *Dolichos biflorus* agglutinin; FGR, fetal growth restriction; HLA, human leukocyte antigen; KIR, killer cell Ig-like receptor; MMP, matrix metalloproteinase; uNK, uterine natural killer.

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is evidence to suggest that the majority of uNK cell precursors originate from outside the uterus. In the early 2000s, Croy and colleagues were unable to detect differentiated uNK cells in NK cell-deficient animals engrafted with parts of wild type uteri, suggesting that uNK cell precursors are extra-uterine (Chantakru *et al.*, 2002). More recently, analysis of changes in uNK cell surface markers during pregnancy suggest that uNK cells endogenous to the uterus decline between gestation day (gd) 0 and gd6, perhaps because their cytotoxicity would be lethal to the foreign conceptus (Takashima *et al.*, 2013). Therefore, the endogenous uNK cell population may be less critical for the maintenance of pregnancy than uNK cells that differentiated from extra-uterine precursors.

Macrophage-derived interleukin-15 (IL-15) is a critical regulator of NK and uNK cell differentiation. It was first observed that the time course of IL-15 expression during pregnancy parallels that of uNK cell granule contents (Ye *et al.*, 1996). Soon thereafter, IL-15^{-/-} animals were generated (Kennedy *et al.*, 2000). IL-15^{-/-} females lack uNK cells, MLAPs, and spiral artery remodeling despite birthing litters of average size but slightly below-average weight (Ashkar *et al.*, 2003, Barber and Pollard, 2003). Recent microarray analysis comparing IL-15^{-/-} and IL-15^{+/+} animals did not detect differences in expression levels of genes involved in decidualization (Bany *et al.*, 2012). Therefore, the finding of below-average weight pups born to IL-15^{-/-} dams cannot be explained by differences in decidualization and is likely due to the effect of IL-15 on uteroplacental circulation via stimulation of uNK cell differentiation.

Recent studies offer candidate regulators of IL-15 expression. For example, there is evidence that the transcription factor Runx3 acts together with other transcription factors to promote IL-15 expression (Levanon *et al.*, 2014). However, there are also potentially indirect regulators of IL-15 expression. For example, a conditional knockout (cKO) of bone morphogenetic protein receptor 2 (BMPR2) in the female reproductive system demonstrated decreased IL-15 expression (Nagashima *et al.*, 2013). As may be expected, BMPR2 cKOs lacked uNK cells at implantation sites and exhibited defects in placentation.

In another animal model of placental underperfusion, heme oxygenase-1 (HO-1) heterozygotes and knockouts also downregulated IL-15 expression and exhibited fewer uNK cells in the DB and MLAP (Linke *et al.*, 2013). Similar to BMPR2 cKOs, Hmox1^{+/-} dams demonstrated intrauterine growth restriction (IUGR), suggesting poor uteroplacental circulation. Interestingly, treatment with CO, a byproduct of HO's heme metabolism, elevated uNK cell numbers, promoted spiral artery remodeling, and decreased the incidence of fetal death. However, CO treatment did not elevate IL-15 levels. In an attempt to explain why IL-15 is downregulated in Hmox1^{+/-} and Hmox1^{-/-} animals, the authors suggest that CO may affect the activity of macrophages, a demonstrated source of IL-15 in the pregnant uterus (Ye *et al.*, 1996). If true, this hypothesis could address the well-established, counterintuitive protection that smoking confers against preeclampsia. Altogether, these observations suggest potential indirect mechanisms of uNK cell differentiation via regulation of IL-15 expression.

Direct interactions between uNK and trophoblast cells: KIRs and HLAs

Differentiated uNK cells express activating and inhibitory cell surface receptors. Stimulation of these subtypes by trophoblast-

expressed ligands, for example, determines the degree of uNK cell activity. Interestingly, the proportions of activating and inhibitory receptors may shift in the presence of a foreign pathogen to modify uNK cell activity and promote cytotoxicity (Xu *et al.*, 2013b). Many of these uNK cell surface receptors belong to the killer cell Ig-like receptor (KIR) family. KIR A and B haplotypes preferentially express inhibitory and activating receptors, respectively, and bind to fetal trophoblast-expressed human leukocyte antigen C (HLA-C), a major histocompatibility complex (MHC) type I molecule.

Importantly, Moffett and colleagues demonstrated associations of certain KIR-HLA combinations, specifically KIR AA and HLA-C2, with diseases of pregnancy like preeclampsia and miscarriage (Hiby *et al.*, 2008, Hiby *et al.*, 2004). Binding of fetal HLA-C2 to the inhibitory receptor KIR2DL1 in these women may predispose to preeclampsia and other placental disorders via insufficient uNK cell activation (Hiby *et al.*, 2004). In contrast, KIR B women preferentially express uNK cell activating receptors. In these women, HLA-C2 likely binds to the activating receptor KIR2DS1, protecting women against these diseases by activating uNK cells and stimulating trophoblast invasion (Hiby *et al.*, 2010, Hiby *et al.*, 2004). Recent evidence that KIR2DS1 stimulates secretion of cytokines like granulocyte macrophage colony-stimulating factor (GM-CSF) by uNK cells and trophoblast migration supports this paradigm (Xiong *et al.*, 2013).

While HLA-C has attracted attention for its disease associations, trophoblast cells express HLAs other than HLA-C. Specifically, they also express HLA-E and HLA-G. HLA-E binds to CD94/NKG2A, an inhibitory receptor on uNK cells (King *et al.*, 2000), and HLA-G binds to leukocyte immunoglobulin-like receptors (LILRs) on uNK cells (Apps *et al.*, 2007). However, HLA-G also binds to a KIR, CD158d/KIR2DL4, which is expressed in endosomes, not on the uNK cell surface like other KIRs (Rajagopalan, 2010). Binding of HLA-G to KIR2DL4 activates downstream pathways that confer a senescent phenotype on the uNK cell (Rajagopalan and Long, 2012). Supernatants from KIR2DL4-stimulated uNK cells enhance the permeability and angiogenic capacity of human umbilical vein endothelial cells (HUVECs). It is easy, therefore, to imagine a role for uNK cells in placental vascular remodeling via HLA-G stimulation of KIR2DL4.

Despite the attention paid to the consequences of HLA-KIR interactions, there is likely a role for non-KIR uNK cell receptors that bind to non-HLA ligands. Specifically, the aryl hydrocarbon receptor (AHR) is expressed by DBA⁻ uNK cells and may be important for the proliferation of this oft-ignored uNK cell subset (Felker *et al.*, 2013). While Ahr^{-/-} implantation sites demonstrated wild type levels of total uNK cells, DBA⁺ cells were smaller, and DBA⁻ cells were fewer in number (Felker *et al.*, 2013). Ahr^{-/-} animals also demonstrated insufficient spiral artery remodeling. Similarly, loss of natural cytotoxicity receptors (NCR), expressed by DBA⁺ cells, didn't affect total uNK cell numbers but impaired uNK cell maturation and spiral artery remodeling (Felker *et al.*, 2013). While the ligands for these receptors are unknown, they are clearly playing an important role in uNK cell maturation and activity, highlighting the importance of interactions outside the KIR-HLA axis.

Trophoblast-derived factors affect uNK cell recruitment

uNK cell-trophoblast cell crosstalk extends beyond contacts between cell surface proteins. For example, trophoblast-derived

peptides of the calcitonin (CT)/calcitonin gene-related peptide (CGRP) family are involved in maintaining proper placental perfusion, possibly by communicating with uNK cells. The clearest link between a CGRP family member, uNK cells, and spiral artery remodeling is adrenomedullin (AM).

Levels of AM, an anti-inflammatory vasodilator, are physiologically elevated in normal pregnancy (Gibbons *et al.*, 2007) but altered in adverse pregnancy outcomes (Lenhart and Caron, 2012). Polymorphisms in the AM gene are associated with birth weight, glycemic regulation, and preeclampsia (Lenhart *et al.*, 2013). AM localizes to implantation sites and is expressed by the uterine epithelium and fetal trophoblast cells (Li *et al.*, 2006). Fetal trophoblast cells also express the AM receptor, calcitonin receptor-like receptor (CLR) (Tsatsaris *et al.*, 2002). Interestingly, AM^{-/-} females are less fertile due to diminished uterine receptivity (Li *et al.*, 2008), and pups born to AM^{-/-} females are more likely to demonstrate FGR (Li *et al.*, 2006). Altogether, these data suggest a role for maternal AM in implantation and placentation.

However, AM^{-/-} embryos are more likely to exhibit FGR than AM^{+/-} or AM^{+/+} embryos, suggesting a role for fetal-derived AM in placentation. AM^{-/-} placentas demonstrate fewer uNK cells and retention of vascular smooth muscle cells lining spiral arteries compared to AM^{+/+} placentas (Li *et al.*, 2013). Concordantly, placentas from AM^{hi/hi} pregnant females, a gene-targeted animal model of AM overexpression, exhibit 30% more uNK cells than AM^{+/+} placentas and upregulate cytokine, chemokine, and matrix metalloproteinase (MMP) expression. *In vitro*, uNK cell-conditioned media supplemented with AM promotes apoptosis of vascular smooth muscle cells, supporting the emerging role for uNK cells in spiral artery remodeling (Li *et al.*, 2013). It remains to be seen whether AM-mediated uNK cell recruitment and activity is dosage-dependent.

Like AM, adrenomedullin 2 (AM2), also known as intermedin, is physiologically elevated during pregnancy (Chauhan *et al.*, 2007). Administration of an AM2 antagonist causes FGR, highlighting the importance of AM2 in a healthy pregnancy (Chauhan *et al.*, 2006).

AM2 is expressed by trophoblast cells and stimulates their invasion via the mitogen-activated protein kinase (MAPK) signaling pathway (Chauhan *et al.*, 2011, Havemann *et al.*, 2013). AM2 also dose-dependently stimulates HLA-G expression in trophoblasts cells, possibly stimulating KIR2DL4 signaling on uNK cells that could then acquire a senescent phenotype and increase vascular permeability and angiogenesis (Chauhan *et al.*, 2011, Rajagopalan and Long, 2012). It is appealing to test whether AM2 dosage correlates with uNK cell recruitment to the decidua and consequent spiral artery remodeling.

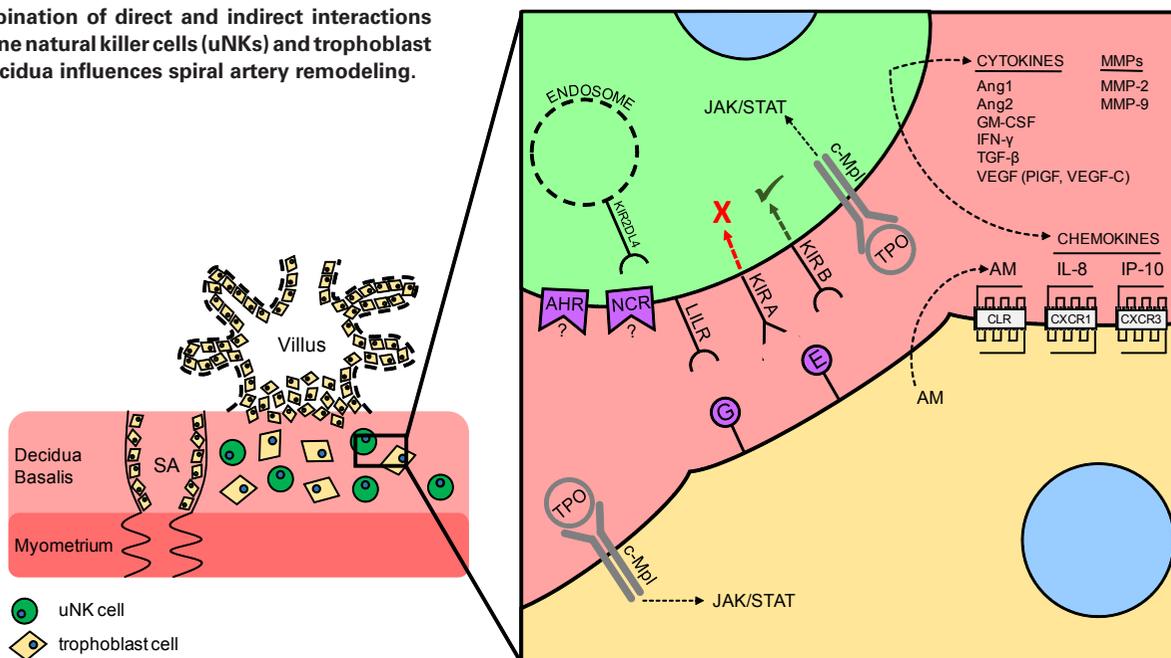
Other CGRP family members also appear to modulate the uteroplacental circulation. For example, CGRP affects blood pressure regulation at the maternal-fetal interface, and its levels are altered in pregnancy-induced hypertension and preeclampsia (Dong *et al.*, 2005, Dong *et al.*, 2004, Fei *et al.*, 2012, Gangula *et al.*, 2003, Knerr *et al.*, 2002). However, CGRPs' effects on uNK cell recruitment and activity remain to be elucidated.

As may be expected, there are trophoblast-derived factors outside the CGRP family that communicate with uNK cells. For example, thrombopoietin (TPO) and its receptor c-Mpl are expressed by uNK cells and trophoblast cells and act in concert to stimulate the proliferation and migration of these cell types via the JAK/STAT pathway (Segeer *et al.*, 2013). As we come to understand more about uNK cell-trophoblast cell crosstalk, we imagine that other trophoblast-derived factors will come to light as important effectors of uNK cell recruitment and activation.

uNK cell-derived factors promote placental vascular remodeling

uNK cells generate an array of angiogenic growth factors, cytokines, and chemokines in different proportions at different times of pregnancy, suggesting a continuous and evolving role for uNK cells as pregnancy progresses. Here, we briefly discuss several examples of uNK cell-derived factors, acknowledging that there

Fig. 1. A combination of direct and indirect interactions between uterine natural killer cells (uNKs) and trophoblast cells in the decidua influences spiral artery remodeling.



are many others we do not address.

Angiopoietin (Ang) 1 and 2, transforming growth factor β (TGF- β), and vascular endothelial growth factor (VEGF) are several examples of uNK cell-secreted angiogenic factors (Lash *et al.*, 2006). Placental growth factor (PIGF) belongs to the VEGF family and has emerged as a potential biomarker for preeclampsia, emphasizing the significance of these angiogenic factors in proper uteroplacental circulation (Chappell *et al.*, 2013, Levine *et al.*, 2004). VEGF-C also belongs to the VEGF family and may assume an additional role in protecting trophoblast cells from uNK cell cytotoxicity (Kalkunte *et al.*, 2009). The concentration of these factors decreases in the decidua as pregnancy progresses, suggesting that they may be irrelevant after spiral artery remodeling concludes (Lash *et al.*, 2006).

Interferon-gamma (IFN- γ) is a uNK cell-secreted cytokine that has arguably attracted the most attention in the literature; it is necessary and sufficient for spiral artery remodeling (Ashkar and Croy, 2001). IFN- γ inhibits trophoblast invasion by promoting apoptosis of trophoblast cells and altering protease levels, keeping trophoblast invasion in check (Lash *et al.*, 2006). Perhaps counterintuitively, uNK cells from preeclamptic women secrete less IFN- γ than uNK cells from normotensive controls (Zhou *et al.*, 2013).

uNK cells also produce a variety of chemokines, including interleukin-8 (IL-8) and interferon-inducible protein-10 (IP-10). Receptors for these two chemokines, CXCR1 and CXCR3, respectively, are expressed on the trophoblast cell surface, substantiating uNK cells' candidacy as potent regulators of trophoblast invasion (Hanna *et al.*, 2006). However, IL-8 and IP-10 presence in uNK cell-conditioned media (CM) isn't different between pregnancies with normal and high uterine artery Doppler resistance indices (RIs) (Wallace *et al.*, 2013). To generate uNK cell CM, the authors plated equal densities of uNK cells from the different pregnancies, which could explain this finding. uNK cell chemokine secretion could very well be equivalent cell to cell, but the two types of pregnancies may have different population sizes of uNK cells. CM from these pregnancies did differ in extracellular signal-regulated kinase (ERK) and Akt pathway activation, which are critical for trophoblast invasion. Therefore, it is possible that uNK cells of these two types of pregnancies differentially express chemokines other than the ones examined in this study.

Finally, uNK cells are also sources of MMPs such as MMP-2 (Naruse *et al.*, 2009). The previously mentioned peptide hormone AM stimulates MMP-9 secretion from uNK cells, triggering spiral artery smooth muscle cell apoptosis (Li *et al.*, 2013). Altogether, this orchestra of signaling molecules coordinates the complex process of spiral artery remodeling to maintain a healthy pregnancy. Intriguingly, angiogenic growth factors and cytokines decrease in concentration when uNK cells and trophoblast cells are co-cultured, though the cell type from which these factors are derived in this co-culture is uncertain (Lash *et al.*, 2011). No doubt additional signals will be identified that will further our understanding of uNK cell-trophoblast interactions.

Conclusions

In summary, the overall immune milieu of the placenta is an important determinant of the health and success of a pregnancy (Arck and Hecher, 2013, Erlebacher, 2013). Perturbations in this complex environment by lipopolysaccharide (LPS)-induced inflammation,

for example, can cause abnormal placental vascular remodeling and phenotypes resembling FGR and preeclampsia (Cotechini *et al.*, 2014). uNK cells dominate this immune landscape during early pregnancy and are important modulators of the maternal-fetal vasculature. Generalized inflammatory changes triggered by obesity, for example, can cause under-recruitment of uNK cells to the decidua, which could explain why obesity elevates a patient's risk of pregnancy complications (Parker *et al.*, 2013).

Certainly, there are other determinants of uNK cell density in the decidua during early pregnancy not discussed here, such as decidual cell-derived cytokines (Lockwood *et al.*, 2013). However, it is likely that there are important effectors of spiral artery remodeling other than uNK cells (Charalambous *et al.*, 2012); several studies suggest that uNK cells may be important only during a small time frame of this process. Ultimately, furthering our understanding of uNK-trophoblast cell interactions and their role in placental vascular remodeling will shed light on placentation disorders like FGR and preeclampsia and may potentially reveal new treatment modalities for these diseases.

References

- APPS, R., GARDNER, L., SHARKEY, A.M., HOLMES, N. and MOFFETT, A. (2007). A homodimeric complex of HLA-G on normal trophoblast cells modulates antigen-presenting cells via LILRB1. *Eur J Immunol* 37: 1924-1937.
- ARCK, P.C. and HECHER, K. (2013). Fetomaternal immune cross-talk and its consequences for maternal and offspring's health. *Nat Med* 19: 548-556.
- ASHKAR, A.A., BLACK, G.P., WEI, Q., HE, H., LIANG, L., HEAD, J.R. and CROY, B.A. (2003). Assessment of requirements for IL-15 and IFN regulatory factors in uterine NK cell differentiation and function during pregnancy. *J Immunol* 171: 2937-2944.
- ASHKAR, A.A. and CROY, B.A. (2001). Functions of uterine natural killer cells are mediated by interferon gamma production during murine pregnancy. *Semin Immunol* 13: 235-241.
- BANY, B.M., SCOTT, C.A. and ECKSTRUM, K.S. (2012). Analysis of uterine gene expression in interleukin-15 knockout mice reveals uterine natural killer cells do not play a major role in decidualization and associated angiogenesis. *Reproduction* 143: 359-375.
- BARBER, E.M. and POLLARD, J.W. (2003). The uterine NK cell population requires IL-15 but these cells are not required for pregnancy nor the resolution of a *Listeria monocytogenes* infection. *J Immunol* 171: 37-46.
- BULMER, J.N., WILLIAMS, P.J. and LASH, G.E. (2010). Immune cells in the placental bed. *Int J Dev Biol* 54: 281-294.
- CHANTAKRU, S., MILLER, C., ROACH, L.E., KUZIEL, W.A., MAEDA, N., WANG, W.C., EVANS, S.S. and CROY, B.A. (2002). Contributions from self-renewal and trafficking to the uterine NK cell population of early pregnancy. *J Immunol* 168: 22-28.
- CHAPPELL, L.C., DUCKWORTH, S., SEED, P.T., GRIFFIN, M., MYERS, J., MACKILLOP, L., SIMPSON, N., WAUGH, J., ANUMBA, D., KENNY, L.C. *et al.*, (2013). Diagnostic accuracy of placental growth factor in women with suspected preeclampsia: a prospective multicenter study. *Circulation* 128: 2121-2131.
- CHARALAMBOUS, F., ELIA, A. and GEORGIADES, P. (2012). Decidual spiral artery remodeling during early post-implantation period in mice: investigation of associations with decidual uNK cells and invasive trophoblast. *Biochem Biophys Res Commun* 417: 847-852.
- CHAUHAN, M., BALAKRISHNAN, M., YALLAMPALLI, U., ENDSLEY, J., HANKINS, G.D., THEILER, R. and YALLAMPALLI, C. (2011). Adrenomedullin 2/intermedin regulates HLA-G in human trophoblasts. *Biol Reprod* 85: 1232-1239.
- CHAUHAN, M., ROSS, G.R., YALLAMPALLI, U. and YALLAMPALLI, C. (2007). Adrenomedullin-2, a novel calcitonin/calcitonin-gene-related peptide family peptide, relaxes rat mesenteric artery: influence of pregnancy. *Endocrinology* 148: 1727-1735.
- CHAUHAN, M., YALLAMPALLI, U., REED, L. and YALLAMPALLI, C. (2006). Adrenomedullin 2 antagonist infusion to rats during midgestation causes fetoplacental growth restriction through apoptosis. *Biol Reprod* 75: 940-947.

- CHEN, Z., ZHANG, J., HATTA, K., LIMA, P.D., YADI, H., COLUCCI, F., YAMADA, A.T. and CROY, B.A. (2012). DBA-lectin reactivity defines mouse uterine natural killer cell subsets with biased gene expression. *Biol Reprod* 87: 81.
- COTECHINI, T., KOMISARENKO, M., SPEROU, A., MACDONALD-GOODFELLOW, S., ADAMS, M.A. and GRAHAM, C.H. (2014). Inflammation in rat pregnancy inhibits spiral artery remodeling leading to fetal growth restriction and features of preeclampsia. *J Exp Med* 211: 165-179.
- DONG, Y.L., GREEN, K.E., VEGIRAGU, S., HANKINS, G.D., MARTIN, E., CHAUHAN, M., THOTA, C. and YALLAMPALLI, C. (2005). Evidence for decreased calcitonin gene-related peptide (CGRP) receptors and compromised responsiveness to CGRP of fetoplacental vessels in preeclamptic pregnancies. *J Clin Endocrinol Metab* 90: 2336-2343.
- DONG, Y.L., VEGIRAJU, S., CHAUHAN, M., GANGULA, P.R., HANKINS, G.D., GOODRUM, L. and YALLAMPALLI, C. (2004). Involvement of calcitonin gene-related peptide in control of human fetoplacental vascular tone. *Am J Physiol Heart Circ Physiol* 286: H230-H239.
- ERLEBACHER, A. (2013). Immunology of the maternal-fetal interface. *Annu Rev Immunol* 31: 387-411.
- FEI, X., HONGXIANG, Z., QI, C. and DAOZHEN, C. (2012). Maternal plasma levels of endothelial dysfunction mediators including AM, CGRP, sICAM-1 and tHcy in pre-eclampsia. *Adv Clin Exp Med* 21: 573-579.
- FELKER, A.M., CHEN, Z., FOSTER, W.G. and CROY, B.A. (2013). Receptors for non-MHC ligands contribute to uterine natural killer cell activation during pregnancy in mice. *Placenta* 34: 757-764.
- GANGULA, P.R., THOTA, C., WIMALAWANSA, S.J., BUKOSKI, R.D. and YALLAMPALLI, C. (2003). Mechanisms involved in calcitonin gene-related Peptide-induced relaxation in pregnant rat uterine artery. *Biol Reprod* 69: 1635-1641.
- GIBBONS, C., DACKOR, R., DUNWORTH, W., FRITZ-SIX, K. and CARON, K.M. (2007). Receptor activity-modifying proteins: RAMPing up adrenomedullin signaling. *Mol Endocrinol* 21: 783-796.
- HANNA, J., GOLDMAN-WOHL, D., HAMANI, Y., AVRAHAM, I., GREENFIELD, C., NATANSON-YARON, S., PRUS, D., COHEN-DANIEL, L., ARNON, T.I., MANAS-TER, I. *et al.*, (2006). Decidual NK cells regulate key developmental processes at the human fetal-maternal interface. *Nat Med* 12: 1065-1074.
- HAVEMANN, D., BALAKRISHNAN, M., BORAHAY, M., THEILER, R., JENNINGS, K., ENDSLEY, J., PHELPS, J., HANKINS, G.D., YALLAMPALLI, C. and CHAUHAN, M. (2013). Intermedin/adrenomedullin 2 is associated with implantation and placentation via trophoblast invasion in human pregnancy. *J Clin Endocrinol Metab* 98: 695-703.
- HIBY, S.E., APPS, R., SHARKEY, A.M., FARRELL, L.E., GARDNER, L., MULDER, A., CLAAS, F.H., WALKER, J.J., REDMAN, C.W., MORGAN, L. *et al.*, (2010). Maternal activating KIRs protect against human reproductive failure mediated by fetal HLA-C2. *J Clin Invest* 120: 4102-4110.
- HIBY, S.E., REGAN, L., LO, W., FARRELL, L., CARRINGTON, M. and MOFFETT, A. (2008). Association of maternal killer-cell immunoglobulin-like receptors and parental HLA-C genotypes with recurrent miscarriage. *Human Reproduction* 23: 972-976.
- HIBY, S.E., WALKER, J.J., O'SHAUGHNESSY, K.M., REDMAN, C.W., CARRINGTON, M., TROWSDALE, J. and MOFFETT, A. (2004). Combinations of maternal KIR and fetal HLA-C genes influence the risk of preeclampsia and reproductive success. *J Exp Med* 200: 957-965.
- KALKUNTE, S.S., MSELLE, T.F., NORRIS, W.E., WIRA, C.R., SENTMAN, C.L. and SHARMA, S. (2009). Vascular endothelial growth factor C facilitates immune tolerance and endovascular activity of human uterine NK cells at the maternal-fetal interface. *J Immunol* 182: 4085-4092.
- KENNEDY, M.K., GLACCUM, M., BROWN, S.N., BUTZ, E.A., VINEY, J.L., EMBERS, M., MATSUKI, N., CHARRIER, K., SEDGER, L., WILLIS, C.R. *et al.*, (2000). Reversible defects in natural killer and memory CD8 T cell lineages in interleukin 15-deficient mice. *J Exp Med* 191: 771-780.
- KING, A., ALLAN, D.S., BOWEN, M., POWIS, S.J., JOSEPH, S., VERMA, S., HIBY, S.E., MCMICHAEL, A.J., LOKE, Y.W. and BRAUD, V.M. (2000). HLA-E is expressed on trophoblast and interacts with CD94/NKG2 receptors on decidual NK cells. *Eur J Immunol* 30: 1623-1631.
- KNERR, I., DACHERT, C., BEINDER, E., METZLER, M., DOTSCHE, J., REPP, R. and RASCHER, W. (2002). Adrenomedullin, calcitonin gene-related peptide and their receptors: evidence for a decreased placental mRNA content in preeclampsia and HELLP syndrome. *Eur J Obstet Gynecol Reprod Biol* 101: 47-53.
- LASH, G.E., NARUSE, K., ROBSON, A., INNES, B.A., SEARLE, R.F., ROBSON, S.C. and BULMER, J.N. (2011). Interaction between uterine natural killer cells and extravillous trophoblast cells: effect on cytokine and angiogenic growth factor production. *Human Reproduction* 26: 2289-2295.
- LASH, G.E., SCHIESSL, B., KIRKLEY, M., INNES, B.A., COOPER, A., SEARLE, R.F., ROBSON, S.C. and BULMER, J.N. (2006). Expression of angiogenic growth factors by uterine natural killer cells during early pregnancy. *J Leukoc Biol* 80: 572-580.
- LENHART, P.M. and CARON, K.M. (2012). Adrenomedullin and pregnancy: perspectives from animal models to humans. *Trends Endocrinol Metab* 23: 524-532.
- LENHART, P.M., NGUYEN, T., WISE, A., CARON, K.M., HERRING, A.H. and STUEBE, A.M. (2013). Adrenomedullin signaling pathway polymorphisms and adverse pregnancy outcomes. *Am J Perinatol* 31: 327-334.
- LEVANON, D., NEGREANU, V., LOTEM, J., BONE, K.R., BRENNER, O., LESH-KOWITZ, D. and GRONER, Y. (2014). Runx3 regulates interleukin-15 dependent natural killer cell activation. *Mol Cell Biol* 34: 1158-1169.
- LEVINE, R.J., MAYNARD, S.E., QIAN, C., LIM, K.H., ENGLAND, L.J., YU, K.F., SCHISTERMAN, E.F., THADHANI, R., SACHS, B.P., EPSTEIN, F.H. *et al.*, (2004). Circulating angiogenic factors and the risk of preeclampsia. *N Engl J Med* 350: 672-683.
- LI, M., SCHWERBROCK, N.M., LENHART, P.M., FRITZ-SIX, K.L., KADMIEL, M., CHRISTINE, K.S., KRAUS, D.M., ESPENSCHIED, S.T., WILLCOCKSON, H.H., MACK, C.P. *et al.*, (2013). Fetal-derived adrenomedullin mediates the innate immune milieu of the placenta. *J Clin Invest* 123: 2408-2420.
- LI, M., WU, Y. and CARON, K.M. (2008). Haploinsufficiency for adrenomedullin reduces pinopodes and diminishes uterine receptivity in mice. *Biol Reprod* 79: 1169-1175.
- LI, M., YEE, D., MAGNUSON, T.R., SMITHIES, O. and CARON, K.M. (2006). Reduced maternal expression of adrenomedullin disrupts fertility, placentation, and fetal growth in mice. *J Clin Invest* 116: 2653-2662.
- LINZKE, N., SCHUMACHER, A., WOJACKI, K., CROY, B.A. and ZENCLUSSEN, A.C. (2013). Carbon monoxide promotes proliferation of uterine natural killer cells and remodeling of spiral arteries in pregnant hypertensive heme oxygenase-1 mutant mice. *Hypertension* 63: 580-588.
- LOCKWOOD, C.J., HUANG, S.J., CHEN, C.P., HUANG, Y., XU, J., FARAMARZI, S., KAYISLI, O., KAYISLI, U., KOOPMAN, L., SMEDTS, D. *et al.*, (2013). Decidual cell regulation of natural killer cell-recruiting chemokines: implications for the pathogenesis and prediction of preeclampsia. *Am J Pathol* 183: 841-856.
- NAGASHIMA, T., LI, Q., CLEMENTI, C., LYDON, J.P., DEMAYO, F.J. and MATZUK, M.M. (2013). BMPR2 is required for postimplantation uterine function and pregnancy maintenance. *J Clin Invest* 123: 2539-2550.
- NARUSE, K., LASH, G.E., INNES, B.A., OTUN, H.A., SEARLE, R.F., ROBSON, S.C. and BULMER, J.N. (2009). Localization of matrix metalloproteinase (MMP)-2, MMP-9 and tissue inhibitors for MMPs (TIMPs) in uterine natural killer cells in early human pregnancy. *Human Reproduction* 24: 553-561.
- PAFFARO, V.A., JR., BIZINOTTO, M.C., JOAZEIRO, P.P. and YAMADA, A.T. (2003). Subset classification of mouse uterine natural killer cells by DBA lectin reactivity. *Placenta* 24: 479-488.
- PARKER, V.J., SOLANO, M.E., ARCK, P.C. and DOUGLAS, A.J. (2013). Diet-induced obesity may affect the uterine immune environment in early-mid pregnancy, reducing NK-cell activity and potentially compromising uterine vascularization. *Int J Obes (Lond)* 38: 766-774.
- RAJAGOPALAN, S. (2010). Endosomal signaling and a novel pathway defined by the natural killer receptor KIR2DL4 (CD158d). *Traffic* 11: 1381-1390.
- RAJAGOPALAN, S. and LONG, E.O. (2012). Cellular senescence induced by CD158d reprograms natural killer cells to promote vascular remodeling. *Proc Natl Acad Sci USA* 109: 20596-20601.
- SEGERER, S.E., MARTIGNONI, F., BOGDAN, A., MULLER, N., KAPP, M., DIETL, J., RIEGER, L. and KAMMERER, U. (2013). Thrombopoietin modulates the proliferation, migration and cytokine profile of decidual cell subsets during early gestation. *Mol Hum Reprod* 19: 361-368.
- SIEWIERA, J., EL COSTA, H., TABIASCO, J., BERREBI, A., CARTRON, G., BOUTEILLER, P. and JABRANE-FERRAT, N. (2013). Human cytomegalovirus infection elicits new decidual natural killer cell effector functions. *PLoS Pathog* 9: e1003257.
- SMITH, S.D., DUNK, C.E., APLIN, J.D., HARRIS, L.K. and JONES, R.L. (2009). Evidence for immune cell involvement in decidual spiral arteriole remodeling in early human pregnancy. *Am J Pathol* 174: 1959-1971.

- TAKASHIMA, A., ISHIKAWA, F., KUWABARA, T., TANAKA, Y., KINOSHITA, T., ITO, M. and KAKIUCHI, T. (2013). Uterine natural killer cells severely decrease in number at gestation day 6 in mice. *Biol Reprod* 89: 101.
- TSATSARIS, V., TARRADE, A., MERVIEL, P., GAREL, J.M., SEGOND, N., JULLIENNE, A. and EVAÏN-BRION, D. (2002). Calcitonin gene-related peptide (CGRP) and CGRP receptor expression at the human implantation site. *J Clin Endocrinol Metab* 87: 4383-4390.
- WALLACE, A.E., HOST, A.J., WHITLEY, G.S. and CARTWRIGHT, J.E. (2013). Decidual natural killer cell interactions with trophoblasts are impaired in pregnancies at increased risk of preeclampsia. *Am J Pathol* 183: 1853-1861.
- XIONG, S., SHARKEY, A.M., KENNEDY, P.R., GARDNER, L., FARRELL, L.E., CHAZARA, O., BAUER, J., HIBY, S.E., COLUCCI, F. and MOFFETT, A. (2013). Maternal uterine NK cell-activating receptor KIR2DS1 enhances placentation. *J Clin Invest* 123: 4264-4272.
- XU, X., FU, Q., ZHANG, Q., ZHAO, M., GAO, Z., LIU, X., LIU, Y. and HU, X. (2013a). Changes of human decidual natural killer cells cocultured with YFP-Toxoplasma gondii: implications for abnormal pregnancy. *Fertil Steril* 99: 427-432.
- XU, X., ZHAO, M., LIU, X., JIANG, Y., ZHANG, H., ZHAI, X., ZHANG, L. and HU, X. (2013b). Toxoplasma gondii infection regulates the balance of activating and inhibitory receptors on decidual natural killer cells. *PLoS One* 8: e55432.
- YE, W., ZHENG, L.M., YOUNG, J.D. and LIU, C.C. (1996). The involvement of interleukin (IL)-15 in regulating the differentiation of granulated metrial gland cells in mouse pregnant uterus. *J Exp Med* 184: 2405-2410.
- ZHANG, J., CHEN, Z., SMITH, G.N. and CROY, B.A. (2011). Natural killer cell-triggered vascular transformation: maternal care before birth? *Cell Mol Immunol* 8: 1-11.
- ZHOU, J., XIAO, X.M. and WU, Y.H. (2013). Expression of interferon-gamma in decidual natural killer cells from women with hypertensive disorder complicating pregnancy. *J Obstet Gynaecol Res* 40: 670-676.

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