

An immunohistochemical analysis of Rab27B distribution in fetal and adult tissue

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ABSTRACT Regulated secretory pathways coordinated by small Rab GTPases are critically involved in intercellular communications. Here, we report the expression and localization of Rab27B in developing and differentiated epithelial human tissues by immunohistochemistry. Rab27B is poorly expressed in fetal tissues suggesting that several developmental mechanisms involved in epithelial differentiation and functions are mediated by other secretory Rab GTPases, such as Rab27A or Rab3 family members. In adult tissues, Rab27B is expressed in a wide variety of differentiated secretory epithelial cells, including those lining the salivary gland, gastrointestinal, mammary and prostate tracts. The complex pattern of Rab27B expression indicates that dysregulation of Rab27B-mediated secretion may have profound implications for disease pathology.

KEY WORDS: *development, small GTPase, exocytosis, secretory Rab, Rab27*

Secretion is a fundamental biological activity of all eukaryotic cells by which they release substances in the extracellular space. The process demands the exquisite interplay between a number of different protein classes, secretory vesicles, the plasma membrane and the eliciting calcium signal. Different types of secretory vesicles exist and their function depends from cell type to cell type. Electron density provides an elementary classification in dense core or light core vesicles. Rabs, small GTP-binding proteins, are the major regulators of vesicular transport (Stenmark, 2009). Like other small GTPases, Rab proteins are functioning as molecular switches using an enzymatic cycle of GTP-binding, hydrolysis and dissociation for their activation or inactivation (Vetter and Wittinghofer, 2001). Post-translational addition of one or two geranylgeranyl groups allows the Rab protein to anchor vesicle membranes (Leung *et al.*, 2006) and to interact with different effectors (Grosshans *et al.*, 2006).

Six members of the Rab family are associated with various secretory vesicles and are described as drivers of regulated secretion: Rab27A/B and Rab3A/B/C/D (Fukuda, 2008). Detailed studies in animal models identified Rab27B on pituitary endocrine

granules (Zhao *et al.*, 2002), dense granules in platelets (Barral *et al.*, 2002, Tolmachova *et al.*, 2007) and megakaryocytes (Tiwari *et al.*, 2003), urothelial fusiform vesicles (Chen *et al.*, 2003), parotid and pancreatic acinar zymogen granules (Chen *et al.*, 2004, Imai *et al.*, 2004), mast cell secretory granules (Mizuno *et al.*, 2007), gastric parietal tubular vesicles (Suda *et al.*, 2011), and secretory granules in alveolar epithelial type II and Clara cells (Bolasco *et al.*, 2011). Furthermore, Rab27B is abundantly expressed in stomach acid-secreting parietal cells and mucus cells, as shown in a transgenic mouse model expressing beta-galactosidase under the Rab27B promoter (Gomi *et al.*, 2007). Aberrant implementation of Rab27B-mediated secretion contributes to cancer progression. Rab27B promotes invasive growth and metastasis in estrogen receptor (ER)-positive breast cancer cell lines. Increased expression of Rab27B, but not Rab3D and Rab27A, is associated with poor-prognosis breast tumors (Hendrix *et al.*, 2010b, Hendrix *et al.*, 2010c).

Abbreviations used in this paper: ER, estrogen receptor.

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In this manuscript, we examined the expression status and the cellular localization of Rab27B in fetal and adult normal human tissues and discuss the observations in relation to the presumptive functions of Rab27B and other secretory Rab GTPases.

Results

Rab27B in human fetal tissues

During embryological development, light or dense core secretory vesicles deliver morphogens such as basic fibroblast growth factor and wingless/wnt to form a concentration gradient across the developing tissue. Rab27B protein is detected at low levels in fetal tissues at 17-24 weeks of gestation. Rab27B is barely detectable in the cytoplasm of mucosal gastric epithelial cells, the small intestine and gall bladder (Fig. 1 A,B,D). A heterogeneous Rab27B distribution is observed in the exocrine pancreas (Fig. 1E). Rab27B is also detected in oocytes and testicular Leydig cells (Fig. 1 F,G). Rab27B is not detected in colon, ovarian epithelial cells, respiratory tract epithelia and in smooth, skeletal and cardiac muscle cells (Fig. 1 C,F,H,I and data not shown). The Rab3 subfamily mediates membrane fusion throughout embryogenesis. Neurogenin 3 commits pancreatic progenitors to an islet cell fate, through Rab3B expression. Rab3B is localized to the cytoplasm

in human pancreatic β -cells during fetal and postnatal development (Piper Hanley *et al.*, 2010). Rab3 proteins play two distinct stimulating roles for large dense-core vesicle fusion in embryonic chromaffin cells, by facilitating vesicle biogenesis and stabilizing the primed vesicle state (Schonn *et al.*, 2010). In the sea urchin Rab3 is specifically enriched in regions of the embryo with active secretory roles such as the apical and basal aspects of cleaving blastomeres, indicative of bidirectional secretion into the extraembryonic environment and blastocoel. Elevated levels of Rab3 were also found in the mesodermally derived pigment cells that invade and move within the ectoderm. These cells likely rely on Rab proteins to enable mobility by mediating the secretion of enzymes that break adhesion to neighboring cells and the extracellular matrix (Conner and Wessel, 2001).

Rab27B in human adult tissues

Exocrine activities

In salivary glands, Rab27B is detected in epithelial cells of the serous-type acini and those lining the striated ducts (Fig. 2A). In acinar cells, identified by dense zymogen granules, Rab27B is detected at the epithelial cell-cell contacts. In striated ducts, Rab27B staining is strikingly polarized at the luminal pole. Salivary

gland acinar cells secrete serous saliva containing amylase. Rab27B regulates amylase release from rat parotid acinar cells (Imai *et al.*, 2004). Rab3D has been identified on secretory vesicles in parotid glands (Ohnishi *et al.*, 1996).

The tall columnar epithelial cells of the gall bladder show abundant Rab27B expression concentrated at the luminal surface (Fig. 2B). Rab27B transports fusiform vesicles, which deliver uroplakin proteins to the apical pole of urothelial umbrella cells (Chen *et al.*, 2003). Smooth muscle fibers are Rab27B negative, except for occasionally infiltrated mast cells.

In the acinar cells of the pancreas Rab27B shows a diffuse and granular cytoplasmic localization (Fig. 2C). The pancreatic ducts express luminal Rab27B. This network is connected with the exocytosis of zymogen granules and digestive enzyme secretion (Chen *et al.*, 2004). Rab3D is present on zymogen granules (Ohnishi *et al.*, 1997). The pancreatic endocrine islets show a weak Rab27B staining (not shown). In contrast, the close homologue Rab27A is found associated with insulin-containing secretory granules of pancreatic β -cells. Of note, siRNA-mediated Rab27A depletion impairs exocytosis of insulin granules triggered by glucose (Tolmachova *et al.*, 2004, Waselle *et al.*, 2003, Yi *et al.*, 2002).

Protective mucus barriers

In the alimentary tract and lungs, Rab27B is found highly expressed at the epithelial surface of mucus secreting cells. These cells protect the epithelial surface from mechanical, chemical, enzymatic, and microbial damages.

In the gastric corpus Rab27B is concentrated at the

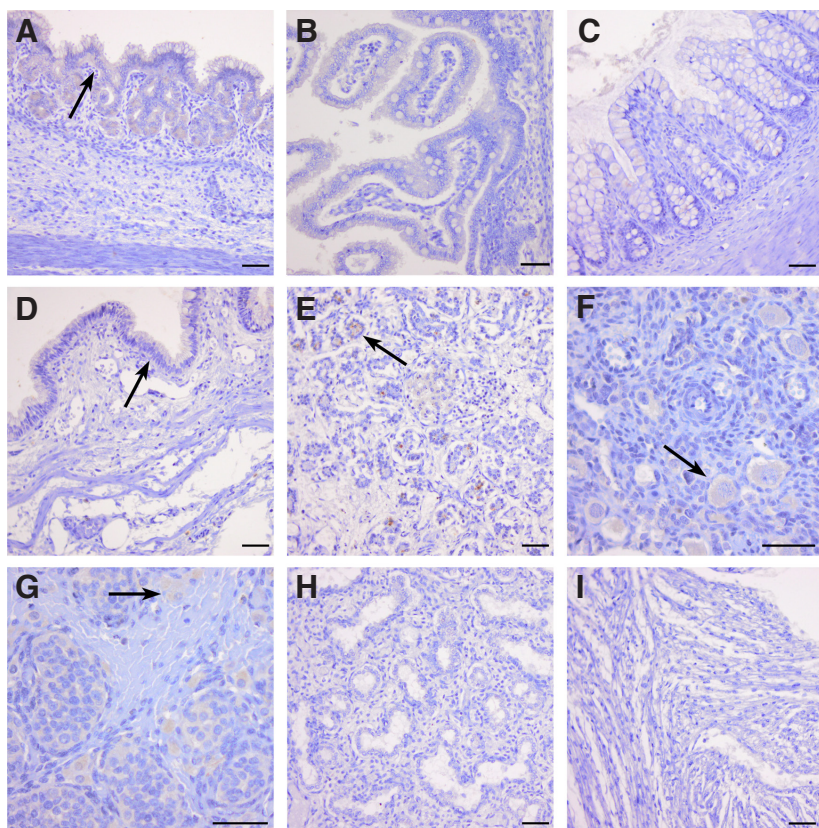


Fig. 1. Immunohistochemical analysis of Rab27B in fetal tissues at 17-24 weeks of gestation. (A) Stomach. (B) Small intestine. (C) Colon. (D) Gall bladder. (E) Pancreas. (F) Ovary. (G) Testis. (H) Lung. (I) Cardiac muscle cells. In (A,D) arrows indicate Rab27B staining in the surface epithelium. In (E) arrow indicates Rab27B positive exocrine cells. Arrows in (F,G) indicate Rab27B expression in oocytes and Leydig cells, respectively. Scale bar: 50 μ m.

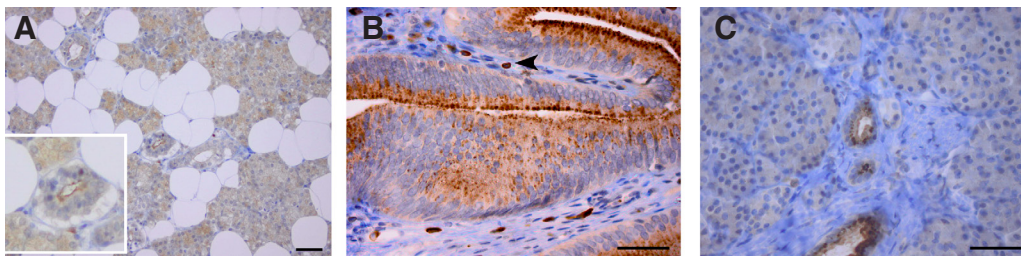


Fig. 2. Rab27B immunostaining in adult tissues with exocrine activity. (A) Salivary gland. Inset: Luminal Rab27B expression in a striated duct. (B) Gall bladder. (C) Pancreas. In (B) arrowhead indicates a mast cell. Scale bar: 50 μ m.

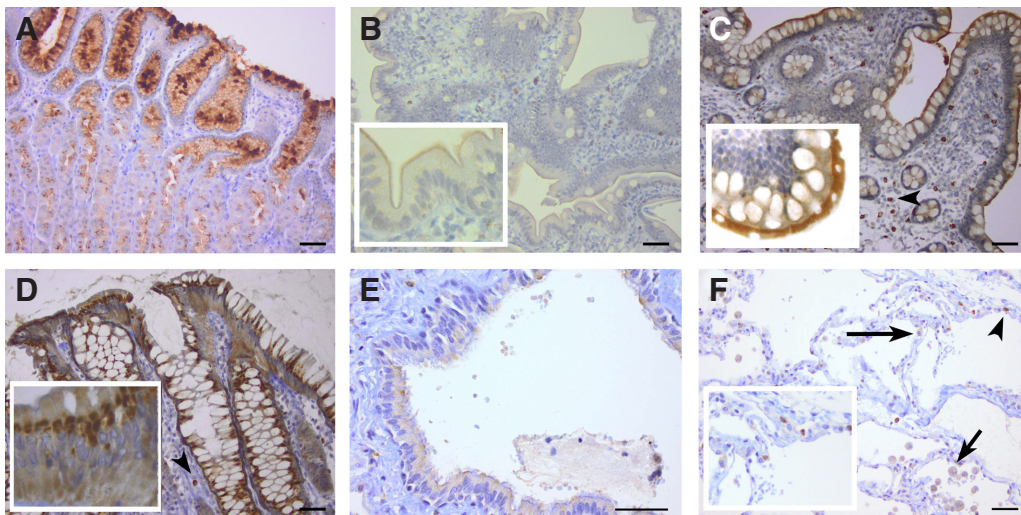


Fig. 3. Rab27B immunostaining in adult tissues with protective mucus barrier. (A) Stomach. (B) Duodenum. (C) Ileum. (D) Colon. (E) Bronchial epithelium. (F) Alveoli. In (F) large arrow, type 2 alveolar pneumocytes; small arrow, macrophage. In (C,D,F) arrowheads indicate mast cells. In (B, C,D) insets indicate Rab27B staining in the surface epithelium. In (F) the inset zooms in on type 2 alveolar pneumocytes and mast cells. Scale bar: 50 μ m.

luminal side of mucus-secreting cells of the surface epithelium and gastric pits (Fig. 3A). In the deep glandular structure consisting of coiled glands, Rab27B localizes at the luminal side. The Rab27B staining pattern in gastric acid-secreting parietal cells varies from a diffuse cytoplasmic localization to concentrated perinuclear signals. Rab27B localizes to tubulovesicular membranes in parietal cells and may play a role in stimulation-induced membrane recruitment of the H⁺/K⁺ATPase proton pump (Suda *et al.*, 2011). Pepsinogen-secreting chief cells exhibit a diffuse Rab27B cytoplasmic staining (Fig. 3A). In the duodenum and ileum epithelia, lined by absorptive cells and interspersed goblet cells, Rab27B staining is detected at the lateral and luminal interfaces (Fig. 3B,C). Interestingly, in the small intestine, Rab27B is less abundant at the base columnar crypts. Absorptive cells and goblet epithelial cells of the colon mucosa demonstrate a strong cytoplasmic Rab27B signal (Fig. 3D). In absorptive cells Rab27B is localized apically according to the basal position of the nuclei, whereas in the goblet cells the Rab27B signal is confined at the basolateral interface, under the apical mucus vacuole. Similarly, Rab27A is present in mucin-secreting cells of the gastrointestinal tract (Tolmachova *et al.*, 2004). In the lamina propria, Rab27B is abundantly detected in the cytoplasm of mast cells.

In the bronchial epithelium Rab27B is detected in the numerous ciliated cells and the scattered goblet cells (Fig. 3E). Both type 2 alveolar pneumocytes and columnar alveolar lining cells involved in surfactant secretions display a cytoplasmic Rab27B expression (Fig. 3F). In contrast, type 1 alveolar pneumocytes and large and flat alveolar lining cells do not express Rab27B protein. Alveolar macrophages and mast cells in the lung parenchyma are Rab27B positive (Fig. 3F). Loss of Rab27B function results in abnormal

lung epithelium structure in mice, characterized by atrophy (Bolasco *et al.*, 2011). Rab3D has been detected in type 2 alveolar pneumocytes, although its precise function remains unknown (van Weeren *et al.*, 2004).

Thrombus formation, allergy and inflammation

In bone marrow, only one type of haematopoietic cell, namely the megakaryocytes, showed abundant and uniform Rab27B staining in the cytoplasm (Fig. 4A,B). Consistently, blood analysis revealed Rab27B expression in platelets but not in myeloid and erythroid cells (data not shown). In platelets, Rab27B expression and function has been described in great detail (Barral *et al.*, 2002, Kondo *et al.*, 2006, Tiwari *et al.*, 2003, Tolmachova *et al.*, 2007). Mature megakaryocytes are converted in proplatelets via the generation of long cytoplasmic extensions coordinated by Rab27B. Proplatelets subsequently generate functional platelets (Tiwari *et al.*, 2003). It is proposed that Rab27B controls the cytoskeletal-mediated transport

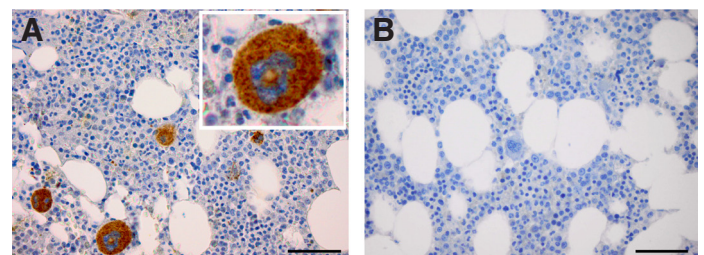


Fig. 4. Rab27B immunostaining in bone marrow (A) and a control slide of bone marrow omitting the primary anti-Rab27B antibody with absence of megakaryocyte staining (B). In (A) the inset indicates a megakaryocyte. Scale bar: 50 μ m.

TABLE 1

Rab27B PROTEIN DISTRIBUTION IN DEVELOPING AND DIFFERENTIATED TISSUES DETERMINED BY IMMUNOHISTOCHEMISTRY

System	Organ	Cell type	Fetal		Adult		
			Localization	Intensity	Localization	Intensity	
Alimentary tract	Salivary	Serous acinar	Nd		Cell-cell contacts	++	
		striated ducts	Nd		Luminal	+++	
	Esophagus	Squamous epithelium		-		-	
		Stomach	Mucus secreting	Diffuse cytoplasmic	+	Luminal	++++
			Chief cells		-	Diffuse cytoplasmic	++
	Small intestine	Parietal cells		-	Diffuse cytoplasmic to concentrated perinuclear	++	
			Absorptive cells	Diffuse cytoplasmic	+	Luminal	++
		Goblet cells	Diffuse cytoplasmic	+	Luminal	++	
		Colon	Absorptive cells		-	Luminal+cytoplasmic	++++
	Goblet cells			-	Luminal+cytoplasmic	++++	
Associated alimentary tract	Liver				-		
	Gall bladder pancreas	Columnar epithelial cells	Diffuse cytoplasmic	+	Luminal	++++	
		Acinar	Diffuse cytoplasmic to concentrated perinuclear	+	Diffuse cytoplasmic	++	
		Pancreatic ducts	Luminal	+	Luminal	+++	
Respiratory	Lung	Endocrine		-		-	
		Ciliated cells		-	Diffuse cytoplasmic	+	
		Goblet cells		-	Diffuse cytoplasmic	+	
		Type I alveolar pneumocytes		-		-	
Haematopoietic	Spleen	Type II alveolar pneumocytes		-	Diffuse cytoplasmic	+	
		Spleen	Nd			-	
		Lymph node	Nd			-	
		Bone marrow	megakaryocytes	Nd		Granular cytoplasmic	++++
Myeloid	Nd				-		
Erythroid	Nd				-		
Reproductive	Mammary gland	Basal myoepithelial	Nd			-	
		Luminal epithelial cells	Nd		Luminal	++++	
	Ovarium	Oocytes	Cytoplasmic	+	Cytoplasmic	+	
		Prostate	Basal cells	Nd			-
	Columnar epithelial cells		Nd		Luminal	++++	
	Testis		Sertoli cells		-		-
		Spermatid		-	Acrosome	++++	
		Myoid cells		-		-	
Soft tissues	Skeletal	Leydig cells	Cytoplasmic	+	Cytoplasmic	++	
		Skeletal muscle cells		-		-	
		Smooth muscle		-		-	
		cardiac	Cardiomyocytes		-		-
All organs		Fibroblasts		-		-	
		Adipocytes		-		-	
		Endothelial cells		-		-	
		Mast cells	Cytoplasmic	+	Granular cytoplasmic	++++	
		macrophages	Cytoplasmic	+	Cytoplasmic	++	

Scales of -, +, ++, +++, +++++ were judged as negative, weakly positive, moderately positive, strongly positive and very strongly positive staining, respectively. Nd, not determined.

and distribution of dense granules during platelet activation and proplatelet formation in megakaryocytes. Platelet aggregation and thrombus formation is supported and enhanced by the release of a number of substances such as serotonin, histamine, ATP, interleukin 1- β and vascular endothelial growth factor from dense granules stored within platelets. The role of Rab27B in allergy and inflammation is highlighted by recent studies showing that Rab27B is localized to histamine and serotonin-containing secretory granules in mast cells, and to azurophilic primary granules in neutrophils, respectively (Johnson *et al.*, 2010, Mizuno *et al.*, 2007). The activation of the high-affinity immunoglobulin E (IgE) receptor induces exocytosis of the granules, resulting in the release of allergy and

inflammatory mediators. Also Rab3 family proteins and Rab27A are present on mast cell secretory granules (Mizuno *et al.*, 2007). Rab27A is detected on primary, secondary and tertiary granules in neutrophils (Herrero-Turrion *et al.*, 2008).

Reproductive tissue

In the mammary gland Rab27B is expressed and concentrated at the luminal surface of the epithelial cell layer (Fig. 5A). The myo-epithelial cell layer and the collagenous fibroblast-rich stroma have no detectable Rab27B protein. In the ovarian stroma no Rab27B expression is detected. Oocytes display a weak cytoplasmic positivity for Rab27B (Fig. 5B).

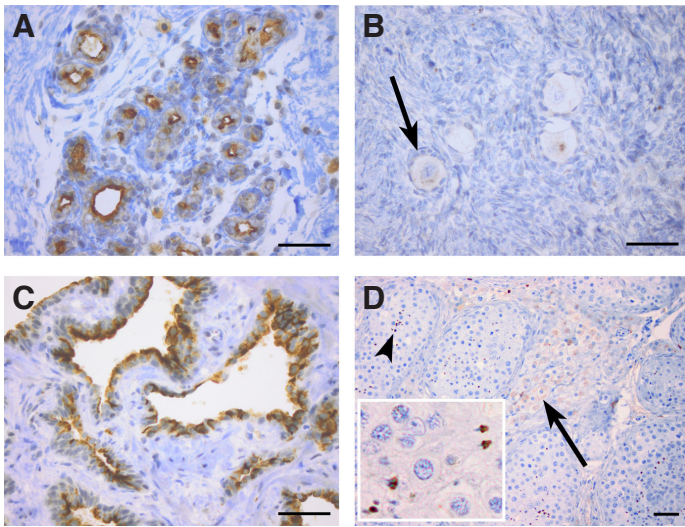


Fig. 5. Analysis of Rab27B protein in (A) mammary gland; (B) ovary (arrow, oocyte); (C) prostate gland and (D) testis (arrow, Leydig cells; arrowhead: spermatids). In (D) the inset zooms in on spermatids. Scale bar: 50 μ m.

The secretory cells of the prostate provide a nutritive and protective medium consisting of proteolytic enzymes, citric acid, acid phosphatase and lipids for the spermatozoa. In the basal cell layer, we observe a weak cytoplasmic Rab27B staining (Fig. 5C) while the stroma is Rab27B negative. In the secretory epithelial cell layer, Rab27B expression is concentrated at the luminal side but also a strong cytoplasmic staining is observed. In the testis Rab27B protein is not detectable in Sertoli cells (Fig. 5D). Spermatogenic cells show a strong Rab27B expression in the acrosome of spermatids. The myoid smooth muscle-like cells that surround the basal lamina of the seminiferous tubules show no Rab27B expression. In the interstitial connective tissue the Leydig cells which secrete testosterone and related androgens, show a weak cytoplasmic Rab27B expression (Fig. 5D). Rab27B is implicated in intramanchette transport during spermiogenesis (Hayasaka *et al.*, 2008).

Discussion

Rab27B does not appear to be critically involved in normal development but is expressed in a broad variety of differentiated secretory cell types.

Fetal tissue array analysis suggests that: 1) The mucosal barrier is absent at the epithelial surface of the fetal bronchus and alimentary tract; 2) The dispersed Rab27B expression in the exocrine cells of the pancreas will likely spread out progressively during gestation to contribute to digestive enzyme secretions. Similar dispersed staining patterns are observed for lipase labeling in the developing human pancreas (Carrere *et al.*, 1992); 3) A critical role for Rab27B in development is not established from our study and other reports since Rab27B-null mice and naturally occurring Rab27B animal mutants exhibit no gross abnormalities in development, behavior and reproduction (Gomi *et al.*, 2007, Tolmachova *et al.*, 2004).

The polarization of Rab27B toward the apical/luminal side of secretory cells coincides with the localization of secretory granules. Our observations are largely consistent with previous analysis of

Rab27B-driven LacZ expression in a mouse model (Gomi *et al.*, 2007). The complex pattern of Rab27B expression (Table 1) indicates that dysregulation of Rab27B-mediated secretion can have profound implications for disease pathology, including cancer. Indeed, increased expression of Rab27B delivers crucial signals for invasion, tumor size, and metastasis of various ER-positive breast cancer cell lines, both in cell culture and in xenograft mouse models (Hendrix *et al.*, 2010b). The presence of Rab27B protein is associated with a low degree of differentiation and lymph node metastasis in ER-positive primary breast cancer. Rab27B is a potential key marker for stratification, prognosis and treatment of early stage ER-positive breast cancers which are more invasive and tend to metastasize more frequently (Hendrix *et al.*, 2010a).

In conclusion, Rab GTPase mediated vesicle exocytosis is efficiently implemented by epithelial cells and bone marrow-derived cells with a secretory function. These data are of (1) biological interest, for the study of cell-type specific Rab27B function, (2) clinical interest, to gain insight in the potential role of Rab27B in different pathologies, and (3) therapeutic interest, to unravel the potential for therapeutic targeting of Rab27B function as suggested in allergy (Mizuno *et al.*, 2007), ER α -positive breast tumors (Hendrix *et al.*, 2010b, Wright *et al.*, 2009) and to predict possible side or toxic effects as a consequence of this therapeutic targeting.

Materials and Methods

Immunohistochemistry

Tissue arrays of normal human fetal tissues were obtained from US Biomax (BE01014 and BE01015; Rockville, MD). De-paraffinization, hydration and antigen-retrieval were performed according to the manufacturer's guidelines. Sections (4 μ m) were cut from non-commercial paraffin-embedded blocks of human adult normal tissues (from each tissue n=3). Sections were fixed in neutral buffered 4% formaldehyde and de-paraffinized with EZ PrepTM Concentrate (Ventana Medical Systems, Tucson, AZ). Subsequently slides were loaded in the Ventana autostainer (Benchmark XT, Ventana Medical Systems) and stained with the Ultraview Universal DAB detection system (Ventana Medical Systems) according to the manufacturer's instructions. Antigen retrieval was performed using EDTA pre-treatment (Ventana Medical Systems). Sections were then incubated with a well-characterized and highly specific anti-Rab27B rabbit polyclonal antibody diluted at 1/500, during 32 min at 37°C (Hendrix *et al.*, 2010b). Nuclei were counterstained with Mayer's haematoxylin. All directly compared images are from slides processed in a single experiment with a matched negative control (purified rabbit immunoglobulin IgG). Four independent observers (two pathologists) performed microscopic evaluation of blind-coded sections.

Conflict of interest

There is no conflict of interest to disclose.

Acknowledgements

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